



## IMPACT OF BLUMEA LACERA ON PROFENOFOS EXPOSED KIDNEY OF WISTAR RAT (RATTUS NOVERGICUS)

Shrutilekha, Singh J. K., Kumari S., and Verma R. K.

P.G. Department of Biotechnology, T. M. Bhagalpur University, Bhagalpur (India)

### ABSTRACT:

*Blumea lacera* has been used as a traditional medicine for anti-inflammatory, antimicrobial, anti-oxidant, anti-helminthic and diuretic. Profenofos is organophosphate compound that generate free radical and damages the cells. In this study, profenofos (50mg/kg bw) was orally administered and 500mg/kg bw *B. lacera* was orally administered. Different kidney function test was performed. It was noticed that after treatment of *B. lacera* concentration of serum creatinine, urea and uric acid was reduced.

**KEY WORLD:** *Blumea lacera*, Profenofos, Wistar rat and Traditional medicine

### INTRODUCTION:

It is very challenging task to fulfill the food stuff to fast growing population (1.252 billions) of India. In order to provide the sufficient food to the growing population, it is necessary to protect the food stuff from pest and spoilage. Farmers use different chemicals like herbicides, insecticides, fungicides, pesticides etc., for crop protection, number of organophosphate have been used frequently from decades in agricultural production, thousands of OPs compounds have been screened and marketed for these purposes (Hassall, 1990; Chirions and Geraud-Pouey, 1996 and Geraud-Pouey, *et al.*, 1997). One of the most frequently used organophosphate is

profenofos. It has adverse effects on many organs like kidney, liver of mice (Akhgari, *et al.*, 2003; Gupta, 2006). It interrupts in the metabolic activity of animals (Aprea, 2000). It also effect the immune system (Neishabouri, *et al.*, 2004), urinary system (Rodrigo, *et al.*, 2001), reproductive system (Joshi, *et al.*, 2003), hematological system (De Blaquiére, *et al.*, 2000), muscles (Pournourmohammadi, *et al.*, 2005) and pancreas (Hagar and Fahmy, 2002). Organophosphate affect the nervous system by disrupting the enzyme acetylcholinesterase hydrolyses acetylcholine into choline and acetic acid (Pandit, *et al.*, 2011 and Yurumez, *et al.*, 2007). Profenofos inhibits the enzyme activity of cholinesterase (Anderson, *et al.*, 1977). Its exposure may cause hepatocellular injury, tissue vacuolization, hemorrhage and hyperplasia of Kupffer cells in the liver (Gomes, *et*

**Corresponding Author** : J. K. Singh  
**E-mail** : jivkant@gmail.com  
**Date of Acceptance** : 20.09.2014  
**Date of Publication** : 30.10.2014

*al.*, 1999). It also leads to swelling of Bowman's capsules and tubular degeneration in the kidney (Fawzy, *et al.*, 2007). It happened due to reactive oxygen stress in different organ of animal (Lin, *et al.*, 2003). During the screening of herbal medicinal plants one of the most precious commodity traditional plants is *Blumea lacera*. *Blumea lacera* is known to work as bitter, astringent, acrid, thermogenic, errhine, anti-inflammatory, styptic, ophthalmic, digestive, anthelmintic, liver tonic, expectorant, febrifuge, antipyretic, diuretic, deobstruant, and stimulant (Warner, *et al.*, 1996). Essential oil of *Blumea* shows analgesic, hypothermic and tranquilizing activities (Anonymous, 1972). Different product of *Blumea* is also used as a homoeopathic drug (Oudhia, *et al.*, 1998) and useful for the treatment of enuresis, neuralgia, headache, cold borne cough and bleeding piles (Ghosh, 1988). In this study, we are trying to look, whether this *Blumea lacera* can useful to treat kidney dysfunction or not due to excessive exposure of profenofos.

#### **MATERIALS AND EXPERIMENTAL PROTOCOLS:**

##### **Model Animal:**

For this present work, twenty four wistar rats with average body weight ranging from 55-60 gm were housed in P.G. Department of Biotechnology, T. M. Bhagalpur University, Bhagalpur (Bihar), India. Food and water to rat were provided ad libitum, formulated feeds were prepared in the laboratory itself. Animals were housed in colony rooms with 12 hrs

light/dark cycle at  $25 \pm 2^{\circ}\text{C}$ .

##### **Extraction of Plant:**

Fresh and mature leaves of *Blumea lacera* was collected and correctly identified with the help of Herbarium of University Department of Botany, T. M Bhagalpur, Bhagalpur (India) having accession number 1078 dated 28<sup>th</sup> August, 2014. Ethanolic and aqueous extraction from *B. lacera* was prepared by soaking 3gm/100ml powdered plant material overnight into different solvents. Extract was filtered and evaporated by using soxhlet till 6 to 8 hrs. Extract was finally dissolve in distilled water and administered in different groups of rat.

##### **Profenofos:**

Profenofos (50% E.C, Specific gravity 1.34, Trade name; "Carina", PI Industries Ltd.) was purchased from the local market and stock solution (50mg/ml) was prepared.

#### **EXPERIMENTAL PROTOCOLS:**

##### **Treatment Protocol:**

Animals were placed in four groups and each containing 6 rats having body weight 55-60gms. experiment was setup as per following protocols.

Group – I : Normal control (NC); Food and water to rats were given ad libitum for 30 days.

Group – II : Profenofos control (PC); 100 $\mu$ l Profenofos orally administered. Animals were exposed to profenofos @ 50mg/kg b.w at alternate day till 30 days. Group – III : Profenofos exposed *B. lacera* (ethanolic extract) treated: After 30 days

of profenofos exposure, *B. lacera* (ethanolic extract) 500mg/kg b.w was orally administered daily till 70 days of experiment initiation. Group – IV : Profenofos exposed *B. lacera* (aqueous extract) treated: After 30 days of profenofos exposure, *B. lacera* (aqueous extract) 500mg/kg b.w was orally administered daily till 70 days of experiment initiation.

#### Blood Collection:

Blood collection from different groups of rats for biochemical analysis was measured through the retro orbital vein puncture by capillary tube. For each experiment maximum 500µl blood was collected in anti-coagulated blood collection tube or plane vile.

#### Biochemical Analysis:

After the entire treatment protocols were used for biochemical analysis of different groups of rats. Blood was collected by orbital sinus vein puncture (Van, et al., 1993). The kidney function test (KFT) i.e., creatinine, urea and uric acid were measured respectively by standard kit and observance was measured at wavelength 520 nm by the help of semi autoanalyzer.

#### RESULTS AND DISCUSSION:

Profenofos is a toxic organophosphate pesticide which kills the pests and simultaneously reduces the hemoglobin concentration in mice (Singh, et al., 2013). It was also reported by Kumar, et al., (2011) Glomerulous of kidney also damaged by profenofos. According to Warner et al., (1996), *B. lacera* has diuretic and anti-oxidant property. Effect of *B. lacera* on profenofos exposed rat, different bio-

chemical tests of different groups of experimental rat were measured. Kidney function test like creatinine, urea and uric acid was measured. It was analysed that normal control of creatinine urea and uric acid was within the normal range. Profenofos control group had increased concentration of these tests. Both ethanolic and aqueous extract of *B. lacera* decreased the concentration as compared with profenofos control but ethanolic extract is more effective than aqueous extract of *B. lacera* (Table 1).

**Table: - 1** Showing different biochemical parameters of kidney functions tests in different groups of experimental mice (data represent in six replicate).

Different Groups	Creatinine (NR:0.6-1.2mg/dl)	Urea (NR:14-40mg/dl)	Uricacid (NR:3.5-7.2mg/dl)
Normal Control	1.0±0.05	35.0±1.0	6.4±0.6
Profenofos	2.7±0.06	61.0±1.5	9.7±0.5
Ethanolic extract of <i>B. lacera</i>	1.4±0.05	36.5±1.0	6.6±0.5
Aqueous extract of <i>B. lacera</i>	1.5±0.04	38±1.0	7.0±0.5

#### ACKNOWLEDGEMENT:

Authors pay their thanks to HOD, P. G. Department of Biotechnology, T. M. Bhagalpur University, Bhagalpur for providing infrastructure for doing this study.

#### REFERENCES:

1. Akhgari M.; Abdollahi M.; Kebryaezadeh A.; Hosseini R. and Sabzevari O.; Biochemical evidence for free radical induced lipid peroxidation as a mechanism for subchronic toxicity of malathion in blood and liver of rats. *Hum Exp Toxicol.* 2003; 22: 205-11.

2. Andreson R. A.; Aaraas I.; Gaare G. and Fonnum F.; Inhibition of acetyl cholinesterase from different species by organophosphorus compounds, carbamates and methyl sulphonyl flouride. *Genetic Pharmac.* 1977; 8: 331-334.
3. Anonymous, *Phytochemistry* 1972; 11: 18-55.
4. Aprea C.; Strambi M.; Novelli M.T.; Lunghini L. & Bozzi N.; Biologic monitoring of exposure to organophosphorus pesticides in 195 Italian children. *Environ Health Perspect*, 2000; 108 (6): 521-5
5. Chirions D. and Geraud-Pouey F.; Effectors de algunos insecticidas sobre entomofauna. *Interciencia*, 1996; 21: 31-36.
6. De Blaquiére G. E.; Waters L.; Blain P. G. and Williams F. M.; Electrophysiological and biochemical effects of single and multiple doses of the organophosphate diazinon in the mouse. *Toxicol Appl Pharmacol*; 2000; 166: 81- 91
7. Fawzy I.; Iman Z., Hamza A. and Ihab A.; The effect of an Organophosphorous insecticide on the hepatic, renal and pulmonary tissues of mice fetuses. *Egypt J. Med. Lab. Sci.*, 2007; 16(2): 99 -113.
8. Geraud-Pouey F.; Chirinos D. and Miranda T. A.; Side effects of insecticide treatments on melon, cucumis mel L. entofauna. *Rev. Fac. Agrono. (LUZ)*, 1997; 14: 225-232.
9. Ghosh N. C., *Comparative materia medica*. Hannemann Publ. Co. Pvt. Ltd., Calcutta, India. 1988; 855.
10. Gomes J.; Dawodu A. H.; Lloyd O.; Revitt D. M. and Anilal S. V.; Hepatic injury and disturbed amino acid metabolism in mice following prolonged exposure to organophosphorus insecticides. *Hum Exp Toxicol.* 1999; 18(1): 33-37.
11. Hagar H. H. and Fahmy A. H.; A biochemical, histochemical, and ultrastructural evaluation of the effect of dimethoate intoxication on rat pancreas. *Toxicol Lett*; 2002; 133: 161-70.
12. Hassall K. A.; *The biochemistry and uses of insecticides*. 2nd Ed., MacMillan Press Ltd., Houndmills, Basingstoke, Taiwan, 1990; 81.
13. Joshi S. C.; Mathur R.; Gajraj A. and Sharma T.; Influence of methyl parathion on reproductive parameters in male rats. *Environ Toxicol Pharmacol*; 2003; 14: 91-98.
14. Kumar A.; Singh J. K.; Md. Ali; Kumar R.; Kumar A.; Nath A.; Roy A. K.; Roy S. P. and Singh J. K.; Evaluation of Cuminum cyminum and Coriandrum sativum on Profenofos induced nephrotoxicity in Swiss albino mice. *Elixir Appl. Botany*, 2011; 39: 4771-4773.
15. Lin L.; Liu J.; Zhang K. and Chen Y.; An experimental study of the effects of profenofos on antioxidase in rabbits. 2003; 32(5): 434-5.
16. Neishabouri E. Z.; Hassan Z. M.; Azizi E. and Ostad S. N.; *Evaluation of immunotoxicity Toxicology* 2004; 196: 173-79.

17. Oudhia P., Joshi B. S.; and Koshta V. K.; Chhattisgarh ke kleshkarak kharptwaron se (The possibilites of preparing homoeopathic drugs from obnoxious weeds of Chhattisgarh. Abstract: V National Science Conference, Bhartiya Krishi Anusandhan Samittee, JNKVV, Gwalior. 1998.
19. Pandit V.; Seshadri S.; Rao S. N.; Samarasinghe C., Kumar A.; and Valsalan R.; A case of organophosphate poisoning presenting with seizure and unavailable history of parenteral suicide attempt. *J Emerg Trauma Shock*; 4: 132-4.
20. Pournourmohammadi S.; Farzami B.; Ostad S. N.; Azizi E.; and Abdollahi M.; Effects of malathion subchronic exposure on rat skeletal muscle glucose metabolism. *Environ. Toxicol Pharmacol*; 2005; 19: 191-96.
21. Rodrigo L.; Hernandez AF.; Lopez-Caballero J. J.; Gil F.; and Pla A.; Immuno histochemical evidence for the expression and induction of paraoxonase in rat; 2001:
22. Singh J. K.; and Roy. A. K.; Role of Curcumin and Cumin on Hematological Parameters of Profenofos Exposed Mice- Mus Musculus, *IJCPRR*. 2013; 4(4): 120-127.
23. Van herck H., et al., Orbital sinus blood sampling in cats as performed by different animal technician: the influence of technique and expertise. *Laboratory Animals*, 1998; 32: 377– 386.
24. Warner P. K.; Nambiar V. R. K.; and Ramakutty C.; Indian Medicinal Plants. Orient Longman, Chennai (India) 1996: 1: 278-280.
25. Yurumez Y.; Durukan P.; Yavuz Y.; Ikizceli I.; Avsarogullari L.; Ozkan S.; Akdur O.; and Ozademir C.; Acute organophosphate poisoning in university hospital emergency room patients. *Intern Med* . 2007; 46 (13): 965-9.
- toxic action of alloxan on pancreatic islet cells in vitro. *Biochem J*, 1979; 182: 17- 25.

\*\*\*